



## Purposeful scientific research is a basic pillar for the advancement of Iraq



### Determination the optimal conditions for the production of antibiotics from some species of *Streptomyces* bacteria that locally isolated

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#### ABSTRACT

The current study was conducted in the postgraduate laboratories of Education College for Pure Sciences at Tikrit-University for the period from the first of December 2019 until the end of March 2020, The study aimed to isolate and identify *Streptomyces* spp. from soil and optimize cultural growth conditions for Antibacterial Metabolites (including antibiotic) Production. Thirteen soil samples were collected from different locations in Tikrit city. Iraq ,Twenty-eight *Streptomyces* isolates were purified and testing their antibacterial activity using primary screening against pathogenic bacteria by perpendicular streaking method and the result showed that all isolates had antibacterial activity in different spectra, The active *Streptomyces* A11 isolate witch exhibited high activity against pathogenic bacteria was identified using several morphological and biochemical methods. Moreover, 16srRNA gene analysis confirmed that this isolate shbowed 99% identity to *Streptomyces vinaceusdrappus*. Ethyl acetate was used to extract the antibacterial compound from fermentation medium, the optimum growth conditions for antibacterial metabolite production by *Streptomyces* A11 isolates were incubation period in 6 days, pH 8 and temperature at 30°C. the GC-MS analysis revealed 12 chemical compounds that may have biological activity .

**Keywords:** *Streptomyces*, Soil, antibacterial activity, GC-MS.

#### 1. Introduction

Members of the genus *Streptomyces* are one of individual group of Actinomycetes known for their abundant production of antibiotics and numerous biologically active secondary metabolic compounds (1). They are Gram positive, high GC-content (2), they spreads by making thread-like hyphae that enable them to obtain nutrients and when nutrient deficient, *Streptomyces* produce air filaments known as aerial hyphae that split and produce spores witch are resistant to unfavorable conditions and can easily spread to new environments (3).

During the growth stage, *Streptomyces* produce secondary metabolic compounds (3), These metabolic by-products aid vegetative bacterial cells to obtain minerals like iron (siderophore), and enabling it to communication with other species and also protect them from UV light by producing pigments (4). The medical importance of *Streptomyces* is that they possess secondary metabolic pathways that produce more than half of the active compounds, including surfactants, antibiotics, anticancer, antiparasitics, antifungals and antibacterial, as well as immunosuppressant's and herbicides (5).

Secondary metabolites production usually occurs at the stationary phase. The nature of secondary metabolites is genetically determined; However, gene

expression can be affected by environmental condition. As a consequence, secondary metabolism usually occurs by supplementation with inducer, nutrient depletion, decreased growth, or by various environmental signals delivered by different organisms in soil niche. (4), the types and quantities of compounds produced by microorganisms are influenced by Many factors, including the nature of the carbon source, nitrogen, pH and aeration, as well as the temperature and the type of strain (6).

#### 2. Material and methods

##### 2.1. Sample collection and Isolation of *Streptomyces* spp.

Soil samples were collected in sterile polyethylene bags from different locations in Tikrit city. soil sample was taken from a depth of 5-20 cm after removing 2 cm from the top layer of soil to avoid surface contamination(7), samples were then dried in electric air oven at 37°C, for four days with the addition of calcium carbonate CaCO<sub>3</sub> in a ratio of 10:1 w/w, One gram from each sample was weighed and added to 10 ml of sterile distilled water, and a serial dilution was performed ( $10^{-1}$ ,  $10^{-2}$ ,  $10^{-3}$ ,  $10^{-4}$  and  $10^{-5}$ ), 0.1 ml of dilution  $10^{-4}$ ,  $10^{-5}$  was spreading on a plate of yeast extract –tryptone agar medium with the following composition (g/l): trypton ;5; yeast

extract, 3; Agar, 15; pH 7.3. (8), This was followed by incubation in inverted position at 28-30°C in an incubator for 7-14 days. Separate colonies were picked up and re-inoculated on plates of the same agar medium by streaking method to ensure the purity of the selected colonies (9).

### **2.2. Primary screening for antibacterial activity**

The antibacterial activity was investigated by using the perpendicular streak plate method (10) against number of pathogenic bacteria which included species of gram-positive bacteria such as *Staphylococcus aureus*, *Staphylococcus saprophyticus*, *Staphylococcus epidermidis*, *Streptococcus pyogenes*, *Enterococcus faecium*, and species of gram-negative bacteria such as *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Serratia marcescens*, *Pseudomonas putida*, *Enterobacter cloacae*, *Proteus mirabilis*, *Escherichia coli*.

### **2.3. Phenotypic Characterization of selected A11 isolate**

Standard Gram stain was carried out and visualized under a compound microscope at X100. Microscopic features like of the substrate and aerial mycelium were observed by using the slide culture or coverslip culture technique (11)(12). The growth ability, color of both substrate and aerial mycelium and production of diffusible pigment of the selected isolate A11 which showed broad spectrum activity antibacterial activity against pathogenic bacteria was determined using different culture media; Yeast extract-malt extract agar, Inorganic salts-starch agar, glucose-asparagine agar, Tyrosine agar, glycerol-asparagine agar and Nutrient agar, and it was subjected to the following biochemical tests: Urease test, catalase test, citrate utilization test, Starch hydrolysis test, Casein hydrolysis test, Gelatin hydrolysis test, Lethinase test, Lipase test, Blood hemolysis test, Tyrosine hydrolysis test and melanin production. In addition to fermenting different types of sugars as a carbon source (Xylose, Inositol, Raffinose, Rhamnose, glucose, sucrose, fructose, mannitol).

### **2.4. Molecular identification of A11 isolate**

The DNA was extracted using Promega Genomic DNA Purification Kit according to the manufacturer instructions. Amplification of 16S rRNA was carried out using the two primers, 27f (5'-AGAGTTTGATCCTGGCTCAG-3') and 1392r (5'-GACGGGCGGTGTGTAC-3'). (13)(14). The PCR amplification included initial denaturation at 94°C for 5 min, followed by 30 cycles at 94°C for 35 s for denaturing, at 55°C for 60 s for annealing, at 72°C for 60 s of extension and at 72°C for 5 min for complete extension (13). The PCR amplification products with the forward and reverse primers were sent to the Micro gene Laboratory in Korea. The 16S rRNA gene base sequences were determined. The results of sequencing were then identified by using the BLAST program (National Centre for Biotechnology Information).

### **2.5. Fermentation and Extraction of Secondary Metabolites and study its biological activity**

The selected isolate A11 was inoculated into flask contain Glycerol-yeast extract broth (5g glycerol, 25g peptone, 2g yeast extract, 0.1 g of K<sub>2</sub>HPO<sub>4</sub>) (15), and then the flasks were placed in the shaking incubator at 30°C and a rotation speed of 150 rpm for 7 days. After that, bacterial biomass was isolated from the production medium using a centrifuge at 10,000 rpm for 20 minutes and then the purification of the production medium was completed using Millipore filter (Whatman No.1). The cell-free supernatant was extracted with an equal volume of ethyl acetate, 1:1 (v/v) and shaken hard for 1 h to complete the extraction process. The aqueous layer was separated from the solvent layer that contain the antibiotic using a separating funnel, a rotary evaporator was used for evaporating the solvent and obtaining a crude precipitate of the antibiotic (16). Agar diffusion method was used to test the sensitivity of pathogenic bacteria to the antibiotic extract, wells 6 mm in diameter were made on Moller Hinton agar medium which was inoculated with the pathogenic test bacteria and each well was filled with 0.1 ml of the residue obtained after dissolved in DMSO (17)(18), then the dishes were incubated at 37 °C for 24 hours and diameters of the inhibition zones were measured (19).

### **2.6. Optimizing Cultural Conditions for antimicrobial compounds Production**

#### **2.6.1. Effect of Incubation Time**

To study the effect of incubation period, flasks contain 50 ml of broth glycerol-yeast extract medium inoculated with selected isolate and the flasks incubated at 30°C in shaking incubator at 150 rpm for 7 days. After incubation, the metabolites were extracted and antibacterial activity was tested daily starting from the second day reaching to the seven day and inhibition zones were measured by using agar diffusion method to determine the optimum period (20).

#### **2.6.2. Effect of pH**

To study the optimal pH for antibiotic production, flasks contain 50 ml of broth glycerol-yeast extract medium was set to pH values ranged from 4 to 9 using 1N HCl or NaOH. The medium were inoculated with selected isolate and placed in shaking incubated at 30°C and 150 rpm for optimum incubation period 6 days, and after that the metabolites were extracted and antibacterial activity was tested (20).

#### **2.6.3. Effect of Incubation Temperature**

The optimum temperature for antibiotic production was determined by using different incubation temperatures (26, 28, 30, 32, 34 and 36) °C. The media at pH 8 were inoculated with selected isolate and placed in shaking incubator at 150 rpm for 6 days (20).

### **2.7. Identification of bioactive compounds**

The antibacterial compounds of the biologically active crude extracts (extracted by ethyl acetate) were identified by using Gas Chromatography-Mass Spectrometer technique (GC MS). The spectrum of the raw ingredients was compared with the spectrum of the recognized ingredients in the National Institute Standard and Technology (NIST) library from which the compound name was determined for the components of the test materials (21). The analysis was carried out using the GC-MS device model 7820A from Agilent company, and the analysis conditions were as follows:

- Culum: Hp-5ms ultra lneit (30m length x)
- 250 nm diameter x 0.25 um in side diameter
- Injection volume: 1 cup
- Pressure: 11.933 psi
- GC inlet line temperature : 250 C
- Aux heaters temperature : 310 C
- Carrier Gas : He 99.99%
- Injection : Temperature : 250 C
- Injection type : Split less
- Scan range : m/z 50-500
- Oven Program:
- Ramp1: 50 C Hold 1 min., Ramp1: 50 C to 150 C, 5 C/ min
- Ramp2: 150 C to 280 C, 8 C/min
- Time amounted to about 37 min

### 3. Results and Discussion

The development of resistance pathogenic bacteria to antibiotics increased the require to search for new sources of antimicrobial agent to fight these pathogenic bacteria. Secondary metabolites produced by bacteria yet interested, because of highly specific antimicrobial activities and their complicated chemical structures. The soil bacteria especially *Streptomyces* spp. are rich sources for many bioactive natural products.

#### 3.1. Isolation of *Streptomyces* spp. and Primary Screening for their Antibacterial Activities

A total of 28 *Streptomyces* spp. were isolated from soil samples collected from different place in tikrit

province, Iraq. Several studies indicate that the soil niches were a rich in many critical actinomycetes (22)(23). All collected isolates were characterized depends on morphology, color of mycelia and diffusible pigment. The results of the preliminary screening by perpendicular streaking method showed that all isolates had antibacterial activity in different spectra. Several previous studies indicate the isolation of *Streptomyces* spp. from soil with antimicrobial activities at a high rate (24)(25).

In this study, A11 isolate showed the highest inhibitory activity against gram positive and gram negative pathogenic bacteria. So, these talented isolate were selected for the production of antibiotics in liquid cultures.

#### 3.2. Phenotypic Characterization and Molecular identification of A11 isolate

The selected isolate were found as long filamentous gram-positive bacteria when examined by light microscope at X100 magnification after staining with Gram stain. When using the slide culture technique Substrate mycelium and aerial mycelium of selected isolate showed clearly.

The Phenotypic characters like color of substrate and aerial mycelia, production of soluble pigment was determined after growing in different growth media Table( 1), The physiological, biochemical characters and utilization of different carbon sources are illustrated in Table 2. Based on these morphological and biochemical characteristics of A11, it was presumptively classified into the genus *Streptomyces*.

Moreover, the partial 16srRNA gene of the selected isolate A11 was sequencing for successful identification. The selected isolate sequence result was matched with previously reported actinobacterial sequences available in the NCBI databases and the result revealed a 99% similarity with *Streptomyces vinaceusdrappus*. The selected isolate 16srRNA sequence was submitted in Genbank (NCBI) and received accession number as MZ618754.1.

**Table 1: Cultural characteristics of A11 isolate on different culture media**

| Characterization                | Culture Medium |                             |                           |                    |
|---------------------------------|----------------|-----------------------------|---------------------------|--------------------|
|                                 | Growth         | Color of substrate mycelium | Color of aerial mycelium  | diffusible pigment |
| Yeast extract-malt extract agar | good           | yellowish cream             | yellowish cream           | Absent             |
| Inorganic salts-starch agar     | good           | cream-gray                  | white-grey                | Absent             |
| glucose -asparagine agar        | good           | yellowish cream             | cream                     | Absent             |
| Tyrosine agar                   | good           | Orange-yellowish brown      | Powdery white -light gray | Absent             |
| glycerol -asparagine agar       | good           | Orange-light brown          | Powdery white             | Absent             |
| Nutrient agar                   | good           | Cream                       | Powdery white             | Absent             |

**Table 2: Physiological and biochemical characteristics of A11selected isolate**

| Test                      | Result |
|---------------------------|--------|
| Catalase activity         |        |
| Urease activity           | +      |
| Starch hydrolysis         | +      |
| Casein hydrolysis         | +      |
| Gelatin hydrolysis        | -      |
| Citrate utilization       | -      |
| Lethinase activity        | +      |
| Lipase activity           | -      |
| Blood heamolysis          | +      |
| Tyrosine hydrolysis       | +      |
| Melanin production        | -      |
| Carbon source utilization |        |
| Glucose                   | +      |
| Sucrose                   | +      |
| Fructose                  | +      |
| Xylose                    | +      |
| Mannitol                  | +      |
| Arabinose                 | +      |
| Rhamnose                  | +      |
| Inositol                  | +      |
| Raffinose                 | +      |

(+)positive reaction and (-) negative reaction

### 3.3. Antibacterial Activity of A11 isolate Secondary Metabolites

Among the Streptomyces isolates that obtained from soil and were screened primarily for their antibacterial activity, only the more active isolate were submitted to secondary screening for the production of metabolic products (include antibiotics) in a liquid culture method depending on antibacterial activity against 12 indicator pathogenic bacteria by the agar diffusion methods, Ethyl acetate has been widely used for secondary metabolites extraction (26) indicated that the secondary metabolites extracted by used ethyl acetate in a 1:1. ratio showed good activity against pathogenic bacteria than other solvents.

Streptomyces A11 isolate Secondary Metabolites exhibited antimicrobial activities against Gram-

positive and Gram-negative pathogenic bacterial isolates.this means that isolate have broad spectrum antibacterial potential (Table 3), The highest inhibition zone against gram negative bacteria was 21 mm against *K.pneumonia* and the lowest inhibition zone of 13 mm against *Entero.cloacae*, while the highest inhibition zone against gram positive bacteria was 26 mm against *Staph.epidermidis* and the lowest inhibition zone of 16 mm against *Staph. aureus* and *Strep.pyogenes*. These results were agree with the previous study conducted by Ahmed *et al* (27) that reported that *S. vinaceusdrappus* has distinct inhibitory effects against Gram-positive and Gram-negative bacteria including *Staph. aureus* and *E. coli* besides antifungal activity and also antitumor against human colon and liver cell lines; and HCT 116and HepG 2 .

**Table 3: Extraction of secondary metabolite by Ethyl acetate solvents**

| Organic solvents | test bacteria           |                       |                          |                      |                       |                    |               |                     |                            |                          |                       |                  |
|------------------|-------------------------|-----------------------|--------------------------|----------------------|-----------------------|--------------------|---------------|---------------------|----------------------------|--------------------------|-----------------------|------------------|
|                  | <i>K.pneumoniae</i>     | <i>Ser.marcescens</i> | <i>Pseudo.aeruginosa</i> | <i>Pseudo.putida</i> | <i>Entero.cloacae</i> | <i>P.mirabilis</i> | <i>E.coli</i> | <i>Staph.aureus</i> | <i>Staph.saprophyticus</i> | <i>Staph.epidermidis</i> | <i>Strep.pyogenes</i> | <i>E.faecium</i> |
|                  | Zone of inhibition (mm) |                       |                          |                      |                       |                    |               |                     |                            |                          |                       |                  |
| Ethyl acetate    | 21                      | 18                    | 19                       | 20                   | 13                    | 16                 | 14            | 16                  | 25                         | 26                       | 16                    | 20               |

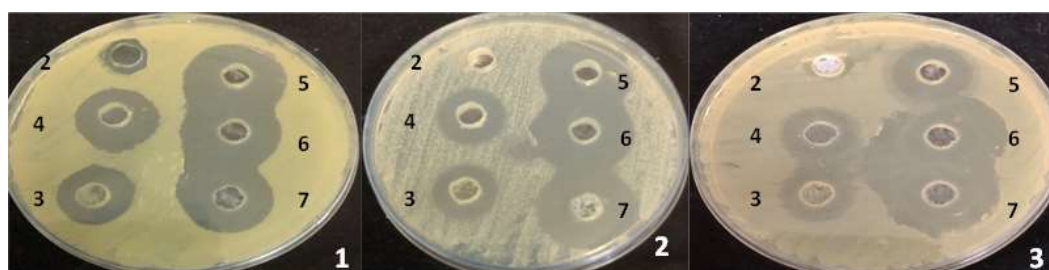
### 3.4. Effect of incubation period

The results showed, as in the table 4, that the highest inhibitory activity against tested pathogenic bacteria by selected isolate A11 was observed at 6 days of incubation, the inhibition zone against pathogenic bacteria ranged between 16mm and 28mm. This is similar to the observation of Thakur *et al* (28) that appropriate incubation period to obtain maximum

antibiotic production from *Streptomyces* spp. 201 was at 6 days later, but differed from earlier reports by BSSN *et al* (29) which indicated that secondary metabolites produced by *S. lavendulocolor* VHB-9 showed the highest antimicrobial activity after 4 days of incubation. The difference could be due to the type of media used in production and varying growth rate of the strains used.

**Table 4: Effect of incubation period**

| incubation period | test bacteria           |                       |                          |                      |                       |                    |               |                     |                            |                          |                       |                  |
|-------------------|-------------------------|-----------------------|--------------------------|----------------------|-----------------------|--------------------|---------------|---------------------|----------------------------|--------------------------|-----------------------|------------------|
|                   | <i>K.pneumoniae</i>     | <i>Ser.marcescens</i> | <i>Pseudo.aeruginosa</i> | <i>Pseudo.putida</i> | <i>Entero.cloacae</i> | <i>P.mirabilis</i> | <i>E.coli</i> | <i>Staph.aureus</i> | <i>Staph.saprophyticus</i> | <i>Staph.epidermidis</i> | <i>Strep.pyogenes</i> | <i>E.faecium</i> |
|                   | Zone of inhibition (mm) |                       |                          |                      |                       |                    |               |                     |                            |                          |                       |                  |
| 2                 | 14                      | 8                     | 11                       | 9                    | 8                     | 11                 | 11            | 8                   | 15                         | 11                       | 8                     | 13               |
| 3                 | 19                      | 10                    | 15                       | 15                   | 10                    | 13                 | 13            | 10                  | 21                         | 21                       | 14                    | 18               |
| 4                 | 20                      | 11                    | 13                       | 16                   | 11                    | 14                 | 13            | 13                  | 21                         | 20                       | 15                    | 20               |
| 5                 | 26                      | 24                    | 16                       | 23                   | 14                    | 15                 | 14            | 19                  | 27                         | 25                       | 17                    | 25               |
| 6                 | 23                      | 25                    | 18                       | 23                   | 15                    | 16                 | 15            | 19                  | 28                         | 28                       | 17                    | 25               |
| 7                 | 21                      | 18                    | 16                       | 19                   | 14                    | 15                 | 14            | 15                  | 25                         | 27                       | 15                    | 21               |



**Fig. 1: effect of different incubation period on the inhibitory activity of the extract prepared from the selected isolate A11**

1- *E.faecium* 2- *Staph.epidermidis* 3- *K.pneumoniae*

### 3.5. Effect of pH

The results revealed that selected isolate A11 produced the antibacterial metabolite (including antibiotic) in different range of pH values (6-9). The highest antibiotic productivity of the selected isolate A11 obtained at pH 8, the diameters of the inhibition zones ranged between 15mm and 28mm (Table 5).

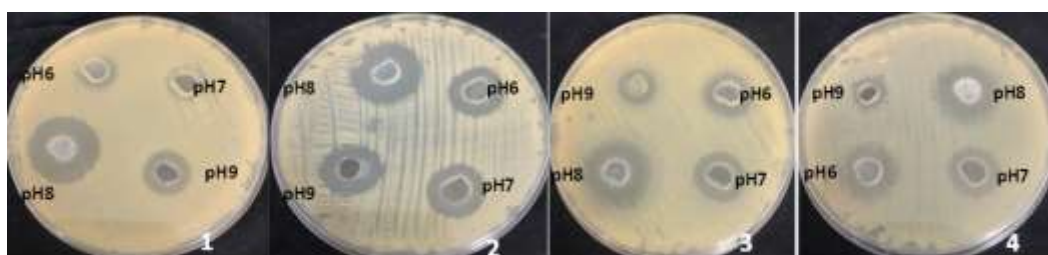
The pH of the culture medium is an important factor affecting the action of cellular enzymes, nutrient ionization, transport across cell membranes, and this will effect on the rate of bacterial growth and the biosynthesis of secondary metabolites.

This result is in agreement with a previous study by Vasavada *et al* (30) which mention that optimum pH for antibiotic production from *Streptomyces sannanensis* RJT-1 was pH 8. Another study by Adeyemo *et al* (31) reported that co-cultured strains of *S. xinghaiensis*-OY62 and *S. rimosus*-OG95 showed excellent antimicrobial activity against different pathogenic bacteria at pH 8.0.

This might be due to the existence of active enzymes for the synthesis of antimicrobial metabolites at this pH, and that hydrogen ion concentration plays a major role in enzyme activity and production of secondary metabolites (32).

**Table 5: Effect of value**

| pH | test bacteria        |                       |                          |                      |                       |                    |               |                     |                            |                          |                       |                  |
|----|----------------------|-----------------------|--------------------------|----------------------|-----------------------|--------------------|---------------|---------------------|----------------------------|--------------------------|-----------------------|------------------|
|    | <i>K.pneumoniae</i>  | <i>Ser.marcescens</i> | <i>Pseudo.aeruginosa</i> | <i>Pseudo.putida</i> | <i>Entero.cloacae</i> | <i>P.mirabilis</i> | <i>E.coli</i> | <i>Staph.aureus</i> | <i>Staph.saprophyticus</i> | <i>Staph.epidermidis</i> | <i>Strep.pyogenes</i> | <i>E.faecium</i> |
|    | inhibition zone (mm) |                       |                          |                      |                       |                    |               |                     |                            |                          |                       |                  |
| 4  | 0                    | 0                     | 0                        | 0                    | 0                     | 0                  | 0             | 0                   | 0                          | 0                        | 0                     | 0                |
| 5  | 0                    | 0                     | 0                        | 0                    | 0                     | 0                  | 0             | 0                   | 0                          | 0                        | 0                     | 0                |
| 6  | 15                   | 14                    | 15                       | 19                   | 9                     | 13                 | 11            | 11                  | 20                         | 21                       | 16                    | 22               |
| 7  | 16                   | 14                    | 17                       | 18                   | 13                    | 13                 | 11            | 11                  | 19                         | 20                       | 11                    | 20               |
| 8  | 25                   | 26                    | 20                       | 26                   | 15                    | 17                 | 16            | 19                  | 27                         | 28                       | 19                    | 26               |
| 9  | 12                   | 18                    | 11                       | 16                   | 8                     | 11                 | 10            | 11                  | 20                         | 19                       | 14                    | 19               |



**Fig. 2: effect of different values of pH on the inhibitory activity of the extract prepared from the selected isolate A11 against pathogenic bacteria**

1- *Ser.marcescens* 2 - *Staph.saprophyticus* 3- *Pseudo.putida* 4-- *K.pneumoniae*

### 3.6. Effect of Temperature

Temperature is one of the factors that have been studied for its effect on the productivity of antibiotics, as it has a direct effect on all metabolic processes, including enzymes activity and active metabolites production (33).The results showed That highest antibiotic yield from selected isolate A11 was

obtained at 30°C and the inhibition zone against pathogenic bacteria ranging between 17mm and 35mm (Table 6), These results agreement with previous report (34)(35). This might be because the metabolic enzymes that enabled the production of antimicrobials were stable at this temperature.

**Table 6: Effect of temperature**

| Temperature | test bacteria        |                       |                          |                      |                       |                    |               |                     |                            |                          |                       |                  |
|-------------|----------------------|-----------------------|--------------------------|----------------------|-----------------------|--------------------|---------------|---------------------|----------------------------|--------------------------|-----------------------|------------------|
|             | <i>K.pneumoniae</i>  | <i>Ser.marcescens</i> | <i>Pseudo.aeruginosa</i> | <i>Pseudo.putida</i> | <i>Entero.cloacae</i> | <i>P.mirabilis</i> | <i>E.coli</i> | <i>Staph.aureus</i> | <i>Staph.saprophyticus</i> | <i>Staph.epidermidis</i> | <i>Strep.pyogenes</i> | <i>E.faecium</i> |
|             | inhibition zone (mm) |                       |                          |                      |                       |                    |               |                     |                            |                          |                       |                  |
| 24          | 0                    | 0                     | 0                        | 0                    | 0                     | 0                  | 0             | 0                   | 0                          | 0                        | 0                     | 0                |
| 26          | 17                   | 15                    | 8                        | 17                   | 14                    | 11                 | 11            | 12                  | 19                         | 26                       | 10                    | 20               |
| 28          | 18                   | 14                    | 10                       | 20                   | 11                    | 14                 | 13            | 15                  | 17                         | 18                       | 12                    | 18               |
| 30          | 25                   | 30                    | 19                       | 26                   | 17                    | 22                 | 23            | 21                  | 29                         | 35                       | 20                    | 28               |
| 32          | 24                   | 28                    | 16                       | 22                   | 15                    | 20                 | 22            | 20                  | 24                         | 31                       | 15                    | 25               |
| 34          | 23                   | 27                    | 14                       | 23                   | 14                    | 20                 | 21            | 19                  | 26                         | 30                       | 14                    | 24               |

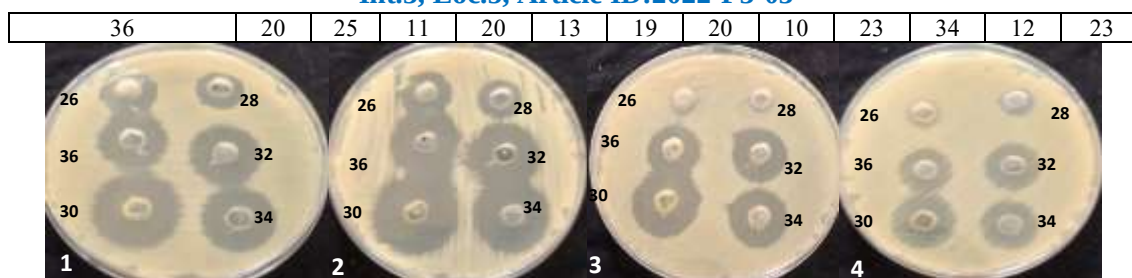


Fig. 3: effect of different incubation temperature on the inhibitory activity of the extract prepared from the selected isolate A11 against pathogenic bacteria

1- *E.faecium* 2-*Staph.saprophyticus* 3-*E.coli* 4-*P.mirabilis*

### 3.7. Identification of bioactive compounds

The GC -MS analysis showed the presence of 19 chemical compounds in the extract of selected isolate A11, Among these compounds are 12 chemical compounds that may have biological activity, namely: 2-Piperidinone(36)(37),Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-(37)(38), Hexadecanoic acid, methyl ester(39)(40), n-Hexadecanoic acid (41), cis-Vaccenic acid (42),2-

Methyl-Z,Z-3,13-octadecadienol(43),oleic acid(44),n-Propyl 11-octadecenoate(45),11-Octadecenoic acid, methyl ester(46),9-Octadecenamamide, (Z)-, 13-Octadecenal (Z)-(44), Heptadecanoic acid, 16-methyl-, methyl ester(47),The presence of the above-mentioned compounds in the extract singly or their synergistic activity with other components may show antimicrobial activity against pathogenic bacteria.

Table 7: Chemical compounds present in the extract of A11isolate using GC-Mass technology

| No. | Compound name                                            | R-Time | Area % |
|-----|----------------------------------------------------------|--------|--------|
| 1   | 2-Piperidinone                                           | 10.459 | 1.58   |
| 2   | 3-Methyl-1,4-diazabicyclo[4.3.0]non-2,5-dione, N-acetyl- | 18.216 | 0.41   |
| 3   | Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-             | 18.853 | 1.62   |
| 4   | Hexadecanoic acid, methyl ester                          | 20.795 | 4.74   |
| 5   | n-Hexadecanoic acid                                      | 21.291 | 0.8    |
| 6   | 2-Methyl-Z,Z-3,13-octadecadienol                         | 22.629 | 0.33   |
| 7   | 9-Octadecenoic acid, methyl ester, (E)-                  | 22.942 | 53.78  |
| 8   | Heptadecanoic acid, 16-methyl-, methyl ester             | 23.179 | 4.1    |
| 9   | Oleic Acid                                               | 23.416 | 19.8   |
| 10  | cis-Vaccenic acid                                        | 23.643 | 2.35   |
| 11  | 9,12-Octadecadienoic acid, methyl ester, (E,E)-          | 23.880 | 2.99   |
| 12  | cis-11-Eicosenoic acid, methyl ester                     | 25.056 | 0.93   |
| 13  | Eicosanoic acid, methyl ester                            | 25.326 | 0.46   |
| 14  | 9-Octadecenamamide, (Z)-                                 | 25.693 | 1.57   |
| 15  | 13-Octadecenal, (Z)                                      | 26.836 | 1.78   |
| 16  | Docosanoic acid, methyl ester                            | 27.365 | 0.42   |
| 17  | Difluoro(methylamino)phosphine sulfide                   | 28.530 | 0.56   |
| 18  | n-Propyl 11-octadecenoate                                | 28.724 | 0.51   |
|     | 11-Octadecenoic acid, methyl ester                       | 28.983 | 1.26   |

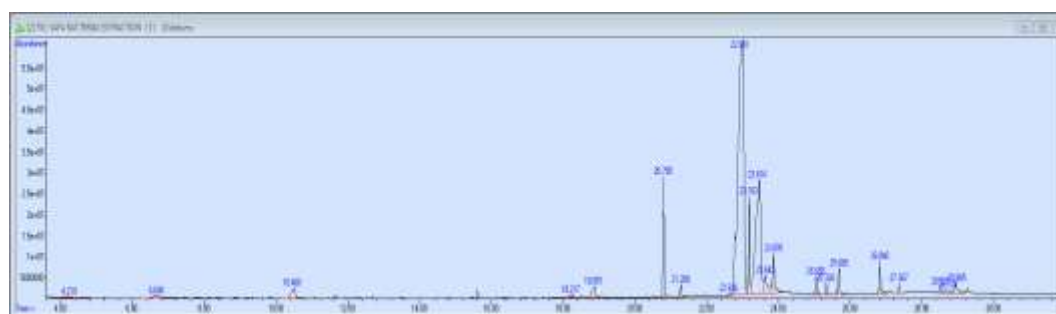


Fig. 4: GC-MS spectrum of antimicrobial compounds produced by A11 isolate.

### Conclusion

Soil Streptomycetes are the main source of bioactive secondary metabolites. In the current study, 13 soil

samples were collected from different location in Tikrit province, Iraq. The active isolate was identified as *S.vinaceusdrappus*, which exhibited broad

spectrum antibacterial activity. Improving culture conditions may effect on the production of secondary metabolites. The GC-MS is one of newer method and the most efficient for identification and

characterization of secondary metabolites. Moreover, this study may be useful for the development of potential antibacterials agents against various pathogens.

## References

- 1- Kong, D., Wang, X., Nie, J., & Niu, G. (2019). Regulation of antibiotic production by signaling molecules in *Streptomyces*. *Frontiers in Microbiology*, 10, 2927.
2. Hopwood, D. A. (2019). Highlights of *Streptomyces* genetics. *Heredity* 123, 23– 32.doi: 10.1038/s41437-019-0196-0
3. Chater, K. F. (2016). Recent advances in understanding *Streptomyces*. *F1000Research*, 5.
4. Bibb M.J. (2005). Regulation of secondary metabolism in *Streptomyces*. *Curr. Opin. Microbiol.*, 8 (2): 208-215
5. Khan, J. A. and Patel, A. S. (2011). Extraction and purification of antibacterial metabolites from *Actinomycetes* spp. isolated from soil samples. *International Journal of Pharmaceutical Research and Development.*, 3 (10): 63-71.
6. Al-Farraj, D. and Mahmoud, A.H.(2020). Antibiotics production in optimized culture condition using low cost substrates from *Streptomyces* sp.AS4 isolated from mangrove soil 7. sediment. *Journal of King Saud University Science.*, 32:1528-1535.
7. Kornwendisch, F and Kutzner, H.(1992) " Family of *Streptomycetaceae* Prokaryotes", Springer-Verlag, New York, pp.921-955.
- 8-. Shirling, E.B. and Gottlieb, D.(1985)"Method for characterization of *streptomyces* species "Journal of systematic Bacteriology, Vol.13, No 54, pp.313-340
9. Williams, S.T. and Davies, F.L.(1995)"Use of antibiotics for selective Isolation and enumeration of actinomycetes in soil" *Journal of General Microbiology*, Vol.38, pp.251-262.
10. Kumar, P.S, Preetam Raj, J.P, Duraipandian, V. and Ignacimuthu, S. (2012). Antibacterial activity of some *Actinomycetes* from Tamil Nadu, India. *Asian Pacific Journal of Tropical Biomedicine.*, 2(12):936-943.
11. Holt, J. ; Krieg, N. ; Sneath, P. ; Staley, J. and Williams, S.T. (1994) "Bergey's manual of Determinative Bacteriology" 9th ed. , Williams and Wilkins Baltimore, pp.605-703
12. Collins, C. H. ; Yates, M. D. and Uttley, H. C. (1988)" Presumptive identification of nocardiasin aclinical laboratory " *Journal of Applied Bacteriology*, Vol.6, No.19, p.55-59.
13. Cotârlet, M., Bahrim, G., Negoita, T. and Stougaard, P. (2010). Comparative study of establishing the efficiency of some methods for chromosomal DNA extraction from cold adapted *Streptomyces*. *Romanian Biotechnological Letters.*, 15(4):5482-5486.
14. Al-sammak, E.G.H.(2017). Phenetic and phylogenetic analysis of *Streptomyces* spp. isolated from hydrocarbons polluted soil. *Al-Nahrain University.*, 11:62-69.
15. Oskay, M., Tamer, U. and Azeri, C., (2004). Antibacterial activity of some actinomycetes isolated from farming soils of Turkey. *Afr. J. Biotechnol.* 3: 441-446.
16. Sahin, N. and Ugur, A.(2003) "Investigation of the antimicrobial activity of some *Streptomyces* Isolates" .*Turkish Journal of Biology Nol.*, No.43 ,pp.79-84 .
17. Augustine S.K., Bhavsar S.P., Baserisallhi M. and Kapadnis B.P. (2004). Isolation, characterization and optimization of antifungal activity of actinomycetes of soil origin. *Ind. J. Exp. Biol.*, 42: 928-932.
18. Khanna, M.; Solanki, R. and Lal, R. (2011). Selective isolation of rare actinomycetes producing novel antimicrobial compounds *Int. J. Adv. Biotech. Res.*, 2 (3) :357-375.
19. Shahrokhi, S.; Bonjar, G. H. and Saadoun, I. (2005). Biological control of potato isolate of *Rhizoctonia solani* by *Streptomyces olivaceus* strain 115. *Biotechnology*;4(2):132-8.
20. Isenberg, H. D. (1992). *Clinical microbiology procedures handbook*. American Society of Microbiology.
21. Chaudhary, R. and Tripathy, A.( 2015). Isolation and identification of bioactive compounds from *Irpex lacteus* wild fleshy fungi. *Journal of Pharmaceutical Sciences and Research.*, 7(7):424-434
22. Savic, M., Bratic, I., and Vasiljevic, B. (2007). *Streptomyces durmitorensis* sp. nov., a producer of an FK506-like immunosuppressant. *Int. J. Syst. Evol. Microbiol.* 57, 2119–2124. doi: 10.1099/ij.s.0.64913-0.
23. Tan, L., Ser, H., Yin, W., Chan, K., Lee, L., and Goh, B. (2015). Investigation of antioxidative and anticancer potentials of *Streptomyces* sp. MUM256 isolated from Malaysia Mangrove Soil. *Front. Microbiol.* 6:1316. doi: 10.3389/fmicb. 2015.01316.
24. Al-Gharani, E. F. H.(2015). Genetic Study of *Streptomycin* Antibiotic Production by *Streptomyces* spp. Bacteria Isolated from Different Soils in Babylon Province. ph.D.Thesis, College of Science, University of Babylon.
25. Al-Asadi, F.M.H.(2016). Extraction and Characterization of Secondary Metabolites produced by *Streptomyces* spp. Isolates in Babylon province. ph.D.Thesis, College of Medicine, University of Babylon.
26. AL-Ghazali, L. H. T. A.( 2017). Production and Characterization of Purified Metabolites from *Streptomyces* Isolate and Study of Antibiotic and

- Anticancer Effects.ph.D.Thesis, College of Science, University of Babylon.
27. Ahmad, M. S., El-Gendy, A. O., Ahmed, R. R., Hassan, H. M., El-Kabbany, H. M., & Merdash, A. G. (2017). Exploring the antimicrobial and antitumor potentials of *Streptomyces* sp. AGM12-1 isolated from Egyptian soil. *Frontiers in microbiology*, 8, 438.
  28. Thakur, D., Bora, T. C., Bordoloi, G. N., & Mazumdar, S. (2009). Influence of nutrition and culturing conditions for optimum growth and antimicrobial metabolite production by *Streptomyces* sp. 201. *Journal de Mycologie Medicale*, 19(3), 161-167.
  29. BSSN, H. B., MUNAGANTI, R. K., MUVVA, V., NARAGANI, K., & INDUPALLI, M. D. (2018). OPTIMIZATION, ISOLATION AND CHARACTERIZATION OF BIOACTIVE COMPOUNDS
  30. Vasavada, S. H. ; Thumar, J. T. and Singh, S. P. (2006). Secretion of a potent antibiotic by salt-tolerant and alkaliphilic actinomycete *Streptomyces sannanensis* strain RJT-1. *Current science*, Vol(91) p.1393-1397.
  31. Adeyemo, O. M., Onilude, A. A., & Babatola, L. J. (2020). Effect of production parameters and inhibitory activity of antimicrobial compounds produced by co-cultured strains of *Streptomyces xinghaiensis*-OY62 and *S. rimosus*-OG95. *Journal of King Saud University-Science*, 32(1), 294-301.
  32. Guimaraes, L.M., Furlan, R.L., Garrido, L.M., Ventura, A., Padilla, G., Facciotti, M.C., 2004. Effect of pH on the production of the antitumor antibiotic rifamycin by *Streptomyces olindensis*. *Biotechnol. Appl. Biochem.* 40, 107–111.
  33. Vijayakumar, R., Panneerselvam, K., Muthukumar, C., Thajuddin, N., Panneerselvam, A., & Saravanamuthu, R. (2012). Optimization of antimicrobial production by a marine actinomycete *Streptomyces afghaniensis* VPTS3-1 isolated from Palk Strait, East Coast of India. *Indian journal of microbiology*, 52(2), 230-239.
  34. Hasaneen, M., El-Sayed, A., & Sabry, S. (2018). Identification, Characterization and Optimized Antimicrobial Production of *Streptomyces thinghirensis* Isolate. *Journal of Agricultural Chemistry and Biotechnology*, 9(12), 263-268.
  35. Moghannem, S. A. (2018). Antibacterial Activity of *Streptomyces Tunisiensis* Mg520500 Isolated from Soil Samples. *The Asia Journal of Applied Microbiology*, 5(1), 1-14.
  36. Kavitha, A., & Savithri, H. S. (2017). Biological significance of marine Actinobacteria of east coast of Andhra Pradesh, India. *Frontiers in microbiology*, 8, 1201.
  37. Al-Maliki, A. D., Ala'a, A. H., & Hameed, M. A. (2021, November). Evaluation of synergistic medicinal efficacy of four alkaloidic compounds isolated from Iraqi *Allium porrum* seeds against *Staphylococcus aureus* bacteria. In *Journal of Physics: Conference Series* (Vol. 2063, No. 1, p. 012026). IOP Publishing.
  38. Kiran, G. S., Priyadharsini, S., Sajayan, A., Ravindran, A., & Selvin, J. (2018). An antibiotic agent pyrrolo [1, 2-a] pyrazine-1, 4-dione, hexahydro isolated from a marine bacteria *Bacillus tequilensis* MSI45 effectively controls multi-drug resistant *Staphylococcus aureus*. *RSC advances*, 8(32), 17837-17846.
  39. Shaaban, M. T., Ghaly, M. F., & Fahmi, S. M. (2021). Antibacterial activities of hexadecanoic acid methyl ester and green-synthesized silver nanoparticles against multidrug-resistant bacteria. *Journal of basic microbiology*, 61(6), 557-568.
  40. Shaaban, M., & El-Mahdy, A. M. (2018). Biosynthesis of Ag, Se, and ZnO nanoparticles with antimicrobial activities against resistant pathogens using waste isolate *Streptomyces enissocaesilis*. *IET nanobiotechnology*, 12(6), 741-747.
  41. Bodoprost, J., & Rosemayer, H. (2007). Analysis of phennacylester derivatives of fatty acids from human skin surface sebum by reversed phase HPTLC: chromatographic mobility as functions of physicochemical properties. *International Journal of Molecular Science*. 8: 1111-1124.
  42. Sani, M. A., Bakar, J., Rahman, R. A., & Abas, F. (2021). Antibacterial composition of bioautographic fractions, characteristics, and stability of *Carica papaya* seed extract. *International Food Research Journal*, 28(3), 443-456.
  43. Adeyemi, M. A., Ekunseitan, D. A., Abiola, S. S., Dipeolu, M. A., Egbeyale, L. T., & Sogunle, O. M. (2017). Phytochemical analysis and GC-MS determination of *Lagenaria breviflora* R. fruit. *International Journal of Pharmacognosy and Phytochemical Research*, 9(7), 1045-1050.
  44. BARTOLOME, A., VILLASENOR, M., YANG, W. 2013. *Bidens pilosa* L. (Asteraceae): Botanical Properties, Traditional Uses, Phytochemistry, and Pharmacology. *Evidence-Based Complementary and Alternative Medicine*, 2013, 1-51.
  45. AL-Amery, S. F., & AlGaraawi, N. L. (2020, December). Phytochemical profile and antifungal activity of stems and leaves methanol extract from the *Juncus maritimus* Linn. *Juncaceae* family against some dermatophytes fungi. In *AIP Conference Proceedings* (Vol. 2290, No. 1, p. 020034). AIP Publishing LLC
  46. Dos Reis, C. M., da Rosa, B. V., da Rosa, G. P., do Carmo, G., Morandini, L. M. B., Ugalde, G. A., ... & Kuhn, R. C. (2019). Antifungal and antibacterial activity of extracts produced from *Diaporthe schini*. *Journal of biotechnology*, 294, 30-37.
  47. Zheng, C. J., Yoo, J. S., Lee, T. G., Cho, H. Y., Kim, Y. H., & Kim, W. G. (2005). Fatty acid synthesis is a target for antibacterial activity of unsaturated fatty acids. *FEBS letters*, 579(23), 5157-5162.

## تحديد الظروف المثلى لإنتاج المضادات الحيوية من بعض أنواع بكتريا *Streptomyces* المعزولة محليا

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### الملخص

أجريت هذه الدراسة في مختبرات الدراسات العليا في قسم علوم الحياة- كلية التربية للعلوم الصرفة-جامعة تكريت لعزل انواع مختلفة من بكتريا *Streptomyces* للفترة من الاول من كانون الاول 2019 ولغاية نهاية شهر اذار 2020، هدفت الدراسة إلى عزل وتشخيص بكتريا *Streptomyces* spp. من التربة وتحديد الظروف المثلى للنمو لإنتاج المؤيضات المضادة للبكتيريا (بما في ذلك المضادات الحيوية). جمعت ثلاثة عشر عينة ترابية من مواقع مختلفة في مدينة تكريت. العراق ، تم تنقية 28 عزلة من بكتريا *Streptomyces* واختبر نشاطها المضاد للبكتيريا بواسطة التحري الأولي ضد البكتيريا المسببة للأمراض بطريقة الخطوط المتعامدة وأظهرت النتائج أن جميع العزلات لها نشاط مضاد للبكتيريا بمديات مختلفة. تم تشخيص العزلة A11 التي اظهرت اعلى فعالية ضد البكتيريا المرضية اعتماداً على الاختبارات الكيموحيوية والصفات المظهرية بالإضافة لذلك أكد تحليل الجين 16srRNA أن هذه العزلة تطابق بنسبة 99 % لـ *Streptomyces vinaceusdrappus*، استخدمت خلاصات الاثيل لاستخلاص المركبات المضادة للبكتيريا الموجودة في وسط التخمر ، كانت ظروف النمو المثلى لإنتاج المؤيضات المضاد للبكتيريا بواسطة العزلة A11 *Streptomyces* هي فترة الحضانة 6 أيام ، ،الاس الهيدروجيني pH8 ودرجة الحرارة 30م. كشف تحليل GC-MS وجود 12 مركب كيميائياً قد يكون لها نشاط بيولوجي.